Cardiopulmonary effects of medetomidine or midazolam in combination with ketamine or tiletamine/zolazepam for the immobilisation of captive cheetahs (Acinonyx jubatus)

G F Stegmann and M Jago

ABSTRACT

Captive cheetahs (Acinonyx jubatus) scheduled for either general health examination or dental surgery were immobilised with combinations of medetomidine-ketamine (K/DET, n = 19), midazolam-ketamine (K/MID, n = 4) or medetomidine-tiletamine-zolazepam (Z/DET, n = 5). Induction time and arterial blood pressure was not statistically significantly (P > 0.05) different between treatment groups. Transient seizures were observed in the K/DET treated animals during induction. Hypertension was present in all groups during anaesthesia with mean (±SD) systolic pressure of 30.7 ± 5.0 kPa for the K/DET group, 27.7 ± 2.7 kPa for the K/MID group, and 33.1 ± 4.6 kPa for the Z/DET group. Heart rate was statistically significantly (P < 0.05) lower in the K/DET group (69 ± 13.2 beats/min) compared to the K/MID group (97 ± 22.6 beats/min), and ventilation rate was statistically significantly (P < 0.05) lower in the K/MID group (15 ± 0.0 breaths/min) compared with the K/DET group (21 ± 4.6). A metabolic acidosis and hypoxia were observed during anaesthesia when breathing air. Oxygen (O2) administration resulted in a statistically significant (P < 0.05) increase in the arterial partial pressure of carbon dioxide (hypercapnoea), arterial partial pressure of O2 and % oxyhaemoglobin saturation.

Key words: Acinonyx jubatus, anaesthesia, cheetah, hypertension, hypoxia, immobilisation, ketamine, medetomidine, seizures, tiletamine, zolazepam.


INTRODUCTION

Reports on the suitability of anaesthetic drugs for immobilisation and anaesthesia in cheetahs (Acinonyx jubatus) are limited. Combinations of xylazine-ketamine or tiletamine-zolazepam, and phen-cyclidine-acepromazine were reported previously. Deem and co-workers reported on the use of tiletamine/zolazepam in combination with medetomidine for immobilisation of cheetahs and observed systemic arterial hypertension during anaesthesia. Hypertension was also reported in other carnivore species after the use of medetomidine. Hypertension is ascribed to be the result of increased peripheral resistance induced by the use of α2-agonists such as medetomidine or xylazine. In wolves the combination of medetomidine and ketamine resulted in prolonged hypertension. In dogs renal failure has also been reported as a cause of hypertension and in humans high blood pressure may aggravate pre-existing chronic renal disease. As kidney disease is known to occur in cheetahs and α2-agonists are known to decrease renal perfusion and increase blood pressure, consideration should be given to the possibility that these drugs may contribute to increased renal morbidity in animals with pre-existing renal disease.

The purpose of this investigation was to examine the cardiopulmonary effects of ketamine/medetomidine (K/DET), tiletamine/zolazepam/medetomidine (Z/DET) and ketamine/midazolam (K/MID) used for immobilisation in captive cheetahs.

MATERIALS AND METHODS

Twenty-eight cheetahs respectively kept either at the De Wildt Cheetah Centre (Pretoria, South Africa) or AfriCat (Otjiwarongo, Namibia) were immobilised for either a general medical examination (AfriCat) or dental surgery (De Wildt). Cheetahs were fasted overnight and water was withheld from the morning prior to immobilisation. Cheetahs were either herded individually into either a crush (De Wildt) and then injected intramuscularly with a pole syringe (De Wildt) or into a small camp (AfriCat) and then darted using a rifle and projectile syringe (Dan-Inject, Sweden). The body weight of the animals was initially estimated for the induction dose to be administered and then accurately weighed after immobilisation for exact dose calculations. Induction time was taken as the time from drug administration until the animals could be handled without reflex movement in response to handling. After immobilisation the anaesthetic depth of the cheetah was monitored by observing response to handling, skeletal muscle tone, heart rate, ventilation rate, mucous membrane colour and rectal temperature at 5-minute intervals. Ambient and rectal temperatures were monitored at 5-minute intervals with the temperature probe from the monitor during the period of immobilisation. When the rectal temperature increased to >39.5 °C the hair coat was sprayed with water, ice packs applied and a electric fan turned on until the temperature decreased to 38 °C. Following recovery from immobilisation, all cheetahs were observed intermittently for a further 24 hours. A full autopsy was conducted on the animal that died.

The cheetahs were immobilised with either K/DET (n = 19), or Z/DET (n = 5), or M/DET (n = 4). At De Wildt the total dose for the K/DET combination varied between 0.2 and 0.4 mg for medetomidine (Domitor, Pfizer) and for ketamine hydrochloride (Anafet, Bayer) varied between 100 and 400 mg. For the Z/DET combination the total dose for ketamine hydrochloride was 400 mg and for midazolam (Domincum, Roche) 15 mg. At AfriCat the total dose for the K/DET combination varied between 0.5 and 1.8 mg for medetomidine and 120 and 180 mg for ketamine. The total dose for tiletamine-zolazepam (Zoletil, Virbac) varied between 60 and...
180 mg. In 3 instances additional ketamine or tiletamine-zolazepam were administered to obtain complete immobilisation (no response to handling) in the cheetah at AfriCat. The choice of drug combination was made by the author without prior knowledge of the individual animal to be immobilised or the procedure to be performed. For dental surgery, anaesthesia was maintained with isoflurane in oxygen. The trachea was intubated and connected to a circle breathing circuit with carbon dioxide absorption with the vapouriser setting at 0.5–1%. Fresh gas flow rate was set at 10 mℓ/kg/min. If cyanosis of the oral mucous membrane was observed when breathing air, the trachea was intubated and the catheter connected to the anaesthetic machine to breathe oxygen. During immobilisation blood volume was maintained with the intravenous administration of a balanced electrolyte solution (Intramed Ringer Lactate Solution) after the placement of a 18 G catheter (Jelco, Johnson & Johnson) in the cephalic vein, and administered at a rate of 10 mℓ/kg/h.

At the end of surgery atipamizole (Antisedan, Pfizer) was administered to antagonise medetomidine in animals where it was used for immobilisation. The dose administered was 0.5 of the volume of medetomidine administered. For recovery, the cheetahs at De Wildt were individually placed in a small pen. At AfriCat the animals were individually placed in crates until recovered (standing without signs of sedation) and thereafter returned to their respective camps.

Arterial blood pressure was measured using a multifunction patient monitor (Critikon Dinamap Plus Vital Signs Monitor). The arterial catheter (20G Jelco, Johnson & Johnson) was connected to a calibrated pressure transducer for measurement of systolic pressure (SYS), diastolic pressure (DIA), mean arterial pressure (MAP) and heart rate (HR). Upon failure to catheterise either the medial saphenous or the femoral artery, an infant-size blood pressure cuff (for 10–19 cm limb circumference) was placed over either the distal humerus or distal femur.

Blood-gas analysis of arterial blood was performed with a blood-gas analyser (Osmetech Opti CCA, Roswell Georgia). Arterial blood was collected anaerobically in 2 mℓ heparinised plastic syringes approximately 15 min after immobilisation. In 3 animals a 2nd sample was taken at the end of dental surgery (sample interval varied between 15 and 30 min). Processing was performed immediately after blood collection. Results were corrected for body temperature.

Data analysis
Data were analysed for normality of distribution and reported as the mean (±SD). The mean values for non-invasive and invasive blood pressure measurements were calculated separately. A general linear model multivariate procedure for multiple dependent variables was used for comparison between treatments. When significant differences were found, Bonferroni’s multiple comparison post hoc test was applied to identify significantly different treatment groups. A 95% confidence interval for significance of difference was accepted (P < 0.05). Owing to the variation in the number of observations over time (length of procedure), only the 1st observation from each variable was used for comparison between treatment groups.

For comparison of the arterial blood-gas variables, and the invasive and non-invasive blood pressure measurements an independent-samples t-test was used. When a 2nd blood-gas sample was collected from the same animal it was only used for comparison of values before and after oxygen administration and not included for calculation of mean values. Data were analysed using a personal computer with SPSS for Windows Ver. 13 software program (SPSS Inc., Chicago, Illinois). The Animal Use and Care Committee of the University of Pretoria approved of this investigation (V064/05).

RESULTS

The mean (±SD) body weight of the cheetahs was 36.8 (±5.2) kg with the range between 26 and 50 kg. The mean induction time for the K/DET group (n = 19) was 9.2 (±3.4) minutes, 11.3 (±1.0) min for the K/MID group (n = 4) and 16.8 (±18.1) min for the Z/DET group (n = 5), and the differences were not statistically significant (P = 0.30). The mean dose for medetomidine was 0.027 mg/kg, for midazolam 0.4 mg/kg, ketamine 6.9 mg/kg, and tiletamine-zolazepam 2.9 mg/kg. Transient seizures were observed in 3 cheetahs immobilised with a high dose of ketamine (400 mg) combined with medetomidine, but not when combined with midazolam.

The mean (±SD) values for ventilation rate, cardiovascular variables, and arterial blood-gas variables are reported in Tables 1 and 2, respectively. Distribution of values was not normal for HR in the Z/DET group (median HR 66). For comparison between treatments, a statistical significant difference (P = 0.034) was found for HR between the K/DET (mean 69 ± 13.2) and K/MID (mean 97 ± 22.6). Differences for HR between K/DET and Z/DET (mean 83 ± 17.3), and K/MID and Z/DET were not statistically significant (P = 0.66, P = 0.17, respectively). Differences for SYS, DIA and MAP between treatments were not statistically significant (P > 0.05). Differences in blood pressure between invasive and non-invasive measurements (Z/DET group) were statistically significant for SYS (P = 0.03), DIA (P = 0.03) and MAP (P = 0.04, Table 1).

The failure rate to catheterise the medial saphenous artery was high, and resulted in placing the catheter in the femoral artery. The failure rate to obtain a blood pressure reading with the cuff was also high.

For ventilation rate a statistical significant difference (P = 0.001) was found between K/DET (mean 21 ± 4.6) and K/MID (mean 15 ± 0). The difference between Z/DET (mean 24.0 ± 7.5) and K/DET or K/MID was not significant (P = 0.92 and P = 0.22, respectively). For arterial blood-gas variables: pH, partial pressure of carbon dioxide (PCO₂), partial pressure of oxygen (PO₂), base excess (BE), plasma bicarbonate (HCO₃), and oxyhaemoglobin saturation (O₂SAT) the differences were not statistically significant (P > 0.05) between the K/DET and Z/DET groups when breathing air (Table 2).

A metabolic acidosis was present in most animals with a pH < 7.40 associated with decreased values for BE and HCO₃. In the K/DET group, administration of oxygen resulted in statistical significant change in the PO₂ (P < 0.03), PCO₂ (P < 0.02), O₂SAT (P < 0.0001) and pH (P < 0.004). The mean rectal temperature was 38.8 °C (±0.9). Environmental temperatures at AfriCat were at maximum 26 °C. High temperatures were experienced at De Wildt (38 °C) and part of the scheduled surgery was cancelled due to the danger of heat stroke.

Recovery was uneventful at AfriCat. At De Wildt, 3 animals from the K/DET group still appeared sedated 24 h after administration of atipamizole. One cheetah from K/MID group died 36 h after anaesthesia. Necropsy examination of this cheetah indicated foreign body aspiration and asphyxia from a commercial dog food meal.

DISCUSSION
The high failure rate to measure arterial blood pressure in the cheetah resulted in the unequal distribution of treatment numbers. It is possible that peripheral vasoconstriction caused by medetomidine ([α₂]-adrenergic agonist) may have contributed towards this failure. The low numbers in the Z/DET (n = 5) and K/MID (n = 4) groups as opposed to the K/DET (n = 19) group may have adversely influenced the power of statistical analysis obtained in this investigation.
The use of either ketamine or tiletamine-zolazepam combined with medetomidine or the combination of ketamine and midazolam had no statistical significant effect ($P < 0.05$) on induction time although a somewhat longer induction time was observed with the Z/DET (16.8 min) combination.

Induction time is of clinical relevance as it should be kept as short as possible as increased locomotion during induction could increase body temperature in hot ambient temperatures. The induction time observed by Deem et al. of 4.3 ± 2 min obtained after tiletamine-zolazepam (1.5 mg/kg) and detomidine (0.03 mg/kg) administration was much shorter than the induction time for the Z/DET group in this investigation although higher doses were used in this investigation. No particular reason for this could be found as the end-point for induction time appeared to be the same. The amount of excitation present in the animal at the time of drug administration is the only factor that may delay induction, and is a possible explanation for the differences in induction times.

The seizures observed in the cheetah induced with a high dose of ketamine (10 mg/kg) have not been reported previously. A dose of 10 mg/kg ketamine in combination with xylazine (1 mg/kg) was recommended by McKenzie and Burroughs. Seizures may occur after ketamine administration in dogs but appear to be benign as no adverse effects have been reported. Medetomidine may decrease the threshold for seizures and could facilitate the onset of seizures in the presence of ketamine. Midazolam increases the seizure threshold in humans and is probably the reason why seizures were not observed with the K/MID group.

Arterial blood pressure was not significantly different ($P < 0.05$) between the anaesthetic combinations (Table 1). Possible hypertensive occurred in all anaesthetic combinations (treatment groups). The blood pressure of conscious cheetahs is not known and it is therefore not possible to objectively interpret the pressures observed during anaesthesia. Hypertension in dogs is defined as systolic/diastolic pressure >23.9/13.3 kPa (180/100 mmHg) though breed differences occur. The MAP of 25.0 ± 3.1 kPa recorded with Z/DET was somewhat less compared with the 28.9 kPa recorded by Deem et al. The pressure obtained with K/MID (22.9 ± 3.5 kPa) compared similar with K/DET (23.5 ± 3.3 kPa) in this investigation. It was accepted that the high blood pressure...
Table 2: Individual and mean ± SD arterial blood-gas values from anaesthetised cheetahs breathing air or oxygen.

<table>
<thead>
<tr>
<th>Anaesthetic</th>
<th>pH units</th>
<th>PCO₂ (kPa)</th>
<th>PO₂ (kPa)</th>
<th>BEI (mmol/l)</th>
<th>HCO₃⁻ (mmol/l)</th>
<th>O₂SAT (%)</th>
<th>Temp (°C)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Zooletil/Medet + air</td>
<td>7.22</td>
<td>6.1</td>
<td>9.2</td>
<td>-7.1</td>
<td>19.8</td>
<td>82</td>
<td>39.5</td>
</tr>
<tr>
<td>+ air</td>
<td>7.35</td>
<td>4.5</td>
<td>10.1</td>
<td>-6.3</td>
<td>18.1</td>
<td>92</td>
<td>37.8</td>
</tr>
<tr>
<td>7.42</td>
<td>3.7</td>
<td>7.3</td>
<td>-5.3</td>
<td>18.0</td>
<td>84</td>
<td>37.8</td>
<td></td>
</tr>
<tr>
<td>7.41</td>
<td>4.1</td>
<td>9.2</td>
<td>-3.9</td>
<td>19</td>
<td>90</td>
<td>38.0</td>
<td></td>
</tr>
<tr>
<td>7.29</td>
<td>3.2</td>
<td>8.2</td>
<td>-12.9</td>
<td>11.4</td>
<td>86</td>
<td>39.3</td>
<td></td>
</tr>
<tr>
<td>Group mean</td>
<td>7.34 ± 0.08</td>
<td>4.3 ± 1.1</td>
<td>8.8 ± 1.1</td>
<td>-7.1 ± 3.4</td>
<td>17.3 ± 3.4</td>
<td>90.0 ± 2.6</td>
<td></td>
</tr>
<tr>
<td>Ket/Medet + air</td>
<td>7.36</td>
<td>4.3</td>
<td>10.1</td>
<td>-6.4</td>
<td>17.5</td>
<td>92</td>
<td>38.0</td>
</tr>
<tr>
<td>+ air</td>
<td>7.33</td>
<td>4.0</td>
<td>11.2</td>
<td>-8.6</td>
<td>14.9</td>
<td>91</td>
<td>40.0</td>
</tr>
<tr>
<td>7.36</td>
<td>4.0</td>
<td>9.4</td>
<td>-7.5</td>
<td>16.2</td>
<td>92</td>
<td>38.0</td>
<td></td>
</tr>
<tr>
<td>7.38</td>
<td>4.0</td>
<td>10.2</td>
<td>-6.3</td>
<td>16.8</td>
<td>91</td>
<td>39.0</td>
<td></td>
</tr>
<tr>
<td>7.38</td>
<td>4.1</td>
<td>8.9</td>
<td>-5.6</td>
<td>17.3</td>
<td>87</td>
<td>39.0</td>
<td></td>
</tr>
<tr>
<td>7.44</td>
<td>3.7</td>
<td>10.6</td>
<td>-3.9</td>
<td>18</td>
<td>93</td>
<td>39.0</td>
<td></td>
</tr>
<tr>
<td>7.42</td>
<td>4.1</td>
<td>8.0</td>
<td>-4.1</td>
<td>19.4</td>
<td>86</td>
<td>37.5</td>
<td></td>
</tr>
<tr>
<td>Group mean</td>
<td>7.38 ± 0.04*</td>
<td>4.0 ± 0.2*</td>
<td>9.8 ± 1.1*</td>
<td>-6.1 ± 1.7*</td>
<td>17.2 ± 1.4*</td>
<td>87.0 ± 4.1*</td>
<td></td>
</tr>
<tr>
<td>Ket/Medet + oxygen</td>
<td>7.27*</td>
<td>6.8</td>
<td>36.0</td>
<td>-4.1</td>
<td>22.9</td>
<td>100</td>
<td>37.4</td>
</tr>
<tr>
<td>+ air</td>
<td>7.21*</td>
<td>6.8</td>
<td>28.6</td>
<td>-7.9</td>
<td>19.7</td>
<td>98</td>
<td>39.9</td>
</tr>
<tr>
<td>7.20*</td>
<td>6.8</td>
<td>32.5</td>
<td>-8.3</td>
<td>19.4</td>
<td>100</td>
<td>39.2</td>
<td></td>
</tr>
<tr>
<td>7.35</td>
<td>7.3</td>
<td>13.0</td>
<td>2.7</td>
<td>29.6</td>
<td>97</td>
<td>38.1</td>
<td></td>
</tr>
<tr>
<td>7.25</td>
<td>6.9</td>
<td>16.2</td>
<td>-5.4</td>
<td>22.0</td>
<td>98</td>
<td>39.4</td>
<td></td>
</tr>
<tr>
<td>7.37</td>
<td>4.7</td>
<td>5.9</td>
<td>-5.1</td>
<td>19.4</td>
<td>64</td>
<td>39.1</td>
<td></td>
</tr>
<tr>
<td>7.23</td>
<td>2.3</td>
<td>12.2</td>
<td>-17.8</td>
<td>7.1</td>
<td>78</td>
<td>40.5</td>
<td></td>
</tr>
<tr>
<td>Group mean*</td>
<td>7.26 ± 0.05*</td>
<td>6.01 ± 2.1*</td>
<td>21.2 ± 10.6*</td>
<td>-6.5 ± 7.4*</td>
<td>20.3 ± 8.2*</td>
<td>94.4 ± 9.2*</td>
<td>38.8 ± 0.9</td>
</tr>
</tbody>
</table>

Ket = ketamine; medet = medetomidine; pCO₂ = carbon dioxide partial pressure; +air = breathing room air; +oxygen = breathing oxygen; pO₂ = oxygen partial pressure; BE = base excess; HCO₃⁻ = bicarbonate; Temp = body temperature; 1 = 1st sample; 2 = 2nd sample from same animal at end of procedure; a, b, c, d = statistically significant difference (P < 0.05); * = excluding duplicates from means.

was the result of peripheral vasoconstriction, a direct effect of medetomidine (a α₂-agonist) on the peripheral blood vessels. Although the hypertension after α₂-agonist administration is reported as transient, the combined administration with ketamine may increase the period of hypertension. In this investigation hypertension was also observed in the cheetahs anaesthetised with ketamine/medetomidine that would indicate that medetomidine is not the only contributor towards the high blood pressure. Other factors that may contribute towards high blood pressure are hypoxia, increased heart rate, and kidney disease. Ketamine is known to increase HR and blood pressure in humans. Hypoxia (pO₂ with values of 8.8 ± 1.1 and 9.8 ± 1.1 kPa, Table 2) was observed in cheetahs breathing air during this investigation.

Increased susceptibility to kidney injury occurs in humans with chronic kidney disease during acute severe elevations in blood pressure. In dogs prolonged high blood pressure may also increase renal injury. Kidney disease is known to occur in captive cheetahs and therefore the possibility should be considered that the administration of drugs that decrease renal perfusion and increase blood pressure could increase renal morbidity during prolonged anaesthesia in animals with pre-existing renal disease.

The clinical significance of high blood pressure in immobilised cheetahs at this stage is not clear but caution should therefore be exercised when administering drugs such as medetomidine or ketamine to cheetahs with possible sub-clinical renal disease. Consideration should also be given to the possibility to antagonise medetomidine as soon as possible when anaesthesia is no longer required, or during prolonged procedures with inhalation anaesthesia to limit the period of peripheral vasoconstriction. The implication is that higher concentrations of inhalation agent will be required for maintenance, and that increased vigilance in monitoring the anaesthetic depth will be necessary when working with potentially dangerous animals.

Of interest to note was that the pressures obtained with the cuff (24.7/14.5 (18.2) kPa) were statistically significant less than the value obtained with direct arterial blood pressure measurement (33.1/21.7 (25.0) kPa) in the Z/DET treatment group (Table 1). The cuff width was within the prescribed 0.55 of limb circumference and therefore could not be the cause of low pressure recordings with the cuff.

A significant difference (P < 0.003) in heart rate was observed between the K/DET 69 ± 13.2 beats/min compared with 97 ± 22.6 in the K/MID group (Table 1). The difference in HR may be explained with the presence of medetomidine. Medetomidine decreases HR and increases peripheral vascular resistance as compared with either midazolam or zoletapam (component of Zoletil) that has minimal effect on heart rate and peripheral vascular resistance.

Hypoxia was observed in cheetahs breathing air (Table 2). The hypoxia was probably the result of ventilatory depression induced by the anaesthetic drugs. The statistically significant lower ventilation rate observed in the K/MID was not unexpected as midazolam is known in humans and goats to result in ventilatory depression. Breathing oxygen when connected to the anaesthetic machine resulted in a statistically significant increase in the PO₂, oxygen saturation, and pCO₂. The increase in pCO₂ is probably the result of loss of the hypoxic ventilatory drive resulting in a decrease in minute ventilation. The metabolic acids present in most animals was probably the result of muscle exertion associated with herding them into a confined space and efforts from the cheetahs to escape. The 2 lowest values for BE (–12.9 and –17.8) were from animals from De Wild and AfriCat and the influence of either of the techniques used during immobilisation could be implicated.

Delayed recovery associated with prolonged sedation was observed in animals immobilised with midazolam, medetomidine and ketamine. Ketamine is metabolised in the liver and excreted through the
kidneys. The principal metabolite of ketamine (as a glucuronide) is active with an increased duration of action⁷. Clinically there was no evidence of disease in these animals before immobilisation with reference to appetite and habitus although the delayed recovery that was observed may indicate reduced renal function. Attempts to evaluate kidney function in cheetahs at De Wildt failed due to a poor correlation between routine kidney function tests such as blood urea nitrogen and creatinine excretion and chronic kidney disease (L. Venter, Faculty of Veterinary Science, University of Pretoria, pers. comm., 2006). The presence of sub-clinical renal disease could thus not be excluded, and may have resulted in delayed renal excretion of ketamine. The cheetah that died from aspiration was immobilised with midazolam and ketamine. Prolonged action after midazolam was also reported in humans in the presence of prolonged excretion of ketamine. The cheetah that died from aspiration was immobilised with midazolam and ketamine. Prolonged action after midazolam was also reported in humans in the presence of renal or liver malfunction¹⁷. Necropsy examination of the cheetah indicated foreign body aspiration and asphyxia from a commercial dog food meal.

In conclusion, the anaesthetic combination did not influence induction time or arterial blood pressure statistically significantly (P > 0.05). Hypertension was observed in all treatment groups. Heart rate was statistically significantly (P < 0.05) lower in the K/DET group compared with the K/MID group, and ventilation rate was statistically significantly (P < 0.05) lower in the K/MID group compared with the K/DET group. A metabolic acidosis, hypoxia (when breathing air) and possible hypertension were observed during anaesthesia. Oxygen administration increased PO₂ and resulted in hypercapnoea during immobilisation.

ACKNOWLEDGEMENTS

The management of De Wildt Cheetah Centre and AfriCat are thanked for their cooperation in making this investigation possible. Mr M. Pieterse from Calicom Trading is thanked for his assistance during processing the blood-gas samples, lending the Osmotech analyser, and donating the Osmotech blood-gas analysis kits used during this investigation.

REFERENCES